

Praktické základy vědecké práce MB130P16

zimní semestr, 0/2, Z, 2 kredity

Přednášející: RNDr. Jan Petrářek, Ph.D.



Agrobacterium-mediated transformation of tobacco BY-2 cells and Arabidopsis cell suspension *Petrářek, J.R. 2002*
Plant Physiology 131, 204-209

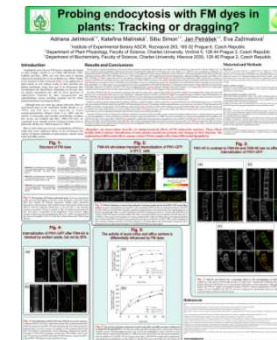
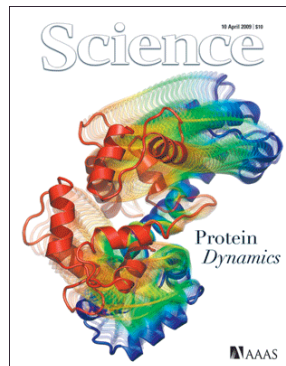
Material and solutions:

- 1) Cell suspension (suspension) cell culture of tobacco BY-2 (30 l in 100 ml spinner flask) or Arabidopsis thaliana cell suspension
- 2) Overlap culture of *Agrobacterium tumefaciens* strains carrying binary vector with your gene construct (GV1001, GV1002, GV1003, GV1004 and others)
- 3) BGJ medium for BY-2 cells (1% Bacto Dextrose, 4.3 g/l Murakige and Skog salts 500MG MOPS, 10 mg/l ascorbic acid, 1 mg/l thiamin, 0.2 mg/l IAA, 2,4-dichlorophenoxyacetic acid, 200 mg/l KH₂PO₄, pH 5.8)
- 4) Arabidopsis thaliana cells (suspension) from your instructor, only Hygromycin and Kanamycin is possible, no PPT is susceptible to bark!
- 5) Plant cell suspension (culture) medium 27°C, kept in BY-2 cells, containing 50 µg/ml Cellvase (Cellulase) and appropriate selection antibiotics
- 6) Glass cell transfer device (pipette) or vortex with 20 µm mesh filter, sterile pipet dishes (6 cm diameter), sterile pipette tips

Procedure:

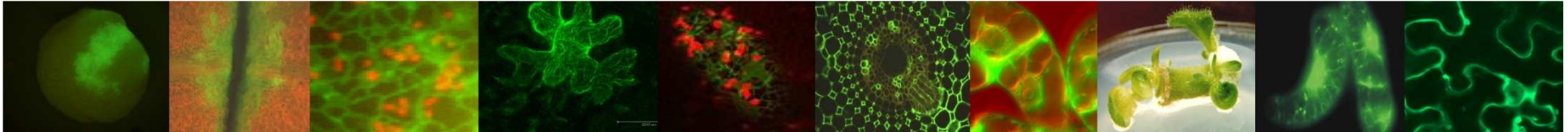
- 1) Filter BY-2 cells and re-suspend them in the same volume of fresh BGJ medium
- 2) Add endotoxin-free water (0.1 ml/ml), using 10 or 20 µm pipette with small tip(s), suck in and out 20 times to remove the wall suspension to help *Agrobacterium* to do its job by producing small vesicles in the medium of BY-2 cells
- 3) Transfer 1 ml of BY-2 suspension into pipette or *Agrobacterium* suspension culture, pipette at least four segments for every construct. The concentration of *Agrobacterium* may range between 20 and 120 (40) ml. Doing this step with only one or only three concentrations. Do not forget to have one plate dish without *Agrobacterium* for control. Wipe with parafilm and incubate 5 days at 27°C in darkness, without changing, it is possible to check with sterile inverted microscope (occasional) preparation of the transformation. It is characterized in that bacteria are attached to cell wall and medium is getting milky, but tobacco cells remain perfectly viable
- 4) Wash cells three times in 50 ml of liquid medium containing high amount of Cellvase (Cellvase, 200 µg/ml) to get rid of the excess of bacteria. Usually all cells from four replicates are mixed and washed together.
- 5) After the last washing step, add 2-3 ml of BY-2 medium with 200 µg/ml Cellvase and transfer this three replicates into plates with appropriate antibiotics (100 µg/ml Kanamycin or 20 µg/ml Hygromycin). Carefully spread cells over the surface of agar plate. This is the crucial step of the whole procedure, cells have to be spread in a thin film in areas enough to allow regeneration.
- 6) Allow regeneration.
- 7) After 48 h grow parafilm and incubate in darkness at 27°C. Antibiotic-resistant clones (0) often appear in 4 weeks of incubation. Cell suspension can be exploited from these colonies. Make the suspension simply by transferring fresh small callus (several millimeters in diameter) into 1.2 l of liquid medium, cut well with pipette. Shake in shaker at 20°C (darkness) or 27°C (Arabidopsis).

Original reference:
An G, (1980) High efficiency transformation of cultured tobacco cells. *Plant Physiol* 70: 559-572



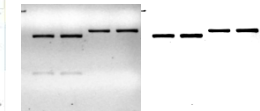
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Co máte šanci se dozvědět a také si vyzkoušet:

- Jak prakticky funguje vědecký tým.
- Jak navrhovat, vést a zpracovávat výsledky experimentů.
- Jak vyhledávat a třídit odbornou literaturu.
- Jak účelně využívat databáze sekvencí.
- Jak napsat bakalářskou a diplomovou práci.
- Jak vyrobit efektní plakát s Vašimi výsledky.
- Jak napsat vědeckou publikaci.



Přednáška/cvičení je určena především pro bakalářské studenty experimentálních oborů, více informací na:
<http://lhr.ueb.cas.cz/petrasek>

